Origin of the Universe – stars, planets, elements

Origin of biorelevant monomers – primordial soup

Complex chemical processes on the way to living systems

Protocells and LUCA

Condensation of aminoacids into peptides



Biochemical condensation of aminoacids into peptides



Nature Publishing Group, www.nature.com/nrg/multimedia



Spontaneous vs. assisted dehydratation



Rode, B. M.; Fitz, D.; Jakschitz, T. Chem. *Biodiversity* **2007**, *4*,2674.

Activating agent	Hydrolysis/ hydration product	$\Delta G^{o\prime}/{ m kJ\ mol^{-1}}$
NH ₂ CONH ₂	$CO_2 + NH_3$	-16^{a}
COS (g)	$CO_2 + H_2S$	-17^{a}
Pyrophosphate	Phosphate	-19^{b}
CO (g)	HCO ₂ H	-16^{a}
HNCO	$CO_2 + NH_3$	-54^{a}
HCN	$HCO_2H + NH_3$	-75^{a}
RCN	$RCO_2H + NH_3$	-80^{c}
NH ₂ CN	Isourea	-83^{d}
HNCNH	Isourea	-97^{d}
HCCH (g)	CH ₃ CHO	-112^{a}

Danger, G.; Plasson, R.; Pascal, R. *Chem. Soc. Rev.* **2012**, *41*, 5416.

Condensation of aminoacids into peptides



Peptide self-replication





Peptide self-replication





K. Severin, D. H. Lee, A. J. Kennan and M. Reza Ghadiri Nature 1997, 389, 706-709

Peptide self-replication



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Nucleotide polymerization

Regioselective formation of 3'-5' phosphodiester bonds between nucleotides





dxline.info/img/new_ail/dna-polymerase_1.jpg







www.neb.com

Products of chemical condensation of nucleotides



- (A) Reaction of an activated mononucleotide (N_{i+1}) with an oligonucleotide (N₁–N_i) to form a 3',5'-phosphodiester (left), 2',5'-phosphodiester (middle), or 5',5'-pyrophosphate linkage (right).
- (B) Typical oligomeric product resulting from chemical condensation of activated mononucleotides

Montmorillonite





B = adenine, guanine, cytosine or uracil



(Top) Structure of ribonucleotide 5'-phosphoimidazolides (left) and ribonucleotide 5'-phosphoro-1-methyladeninium (right). (Bottom) Unit cell of montmorillonite and phosphodiester bond formation within the clay interlayers, as proposed by Ferris and coworkers (right). XH, depicted in blue in the cartoon, is any undifferentiated protic species inside the clay galleries.

Intercalating agents







Template-directed synthesis



Template-directed synthesis





J. Sutherland et al. Nature Chem. 2013, 383-389



Protection of the 2'-OH group of 1-3'P facilitates rapid template-directed 3',5'-ligation after electrophilic phosphate activation. The 3'-OH group of 1-2'P is protected to a lesser extent, such that 1>P is the major product of phosphate activation and slow template-directed 2',5'-ligation follows.

X = leaving group, Y = leaving group generated by electrophilic activation of phosphate oxygen with or without a subsequent nucleophilic displacement

J. Sutherland et al. Nature Chem. 2013, 383-389



Treatment of A3'P (100 mM) with sodium thioacetate 3 (100 mM) and cyanoacetylene 4 (200 mM) in D₂O at neutral pD for 24 hours results in selective acetylation of the 2-OH group. Curly arrows indicate electrophilic activation/acetylation steps. Yields were judged by ¹H NMR integration. Ade = N9-linked adenine.

J. Sutherland et al. Nature Chem. 2013, 383-389



Treatment of A3'P (80 mM) and A2'P (20 mM) as given before results in the exclusive 2-acetylation of the former nucleotide. Partial ¹H NMR spectrum of the reaction products.



Additional electrophiles 6–8 shown to drive the acetylation of ribonucleotides with thioacetate 3. Direct acetylation with 9 is also possible, as is oxidative activation of 3 with ferricyanide 10 to afford ferrocyanide 11 and a dimeric acetylating agent 12. Curly arrows indicate electrophilic activation/acetylation steps.

Chemoselective acetylation of 3'P-oligoribonucleotides expedites templated ligation



Sequences and reaction conditions employed for acetylation (i) and subsequent templated ligation (ii). The acetylation mixture contained 80 mM primer and 50 mM NAI 9; the ligation mixture contained 4 mM primer from the acetylation reaction, 25 mM template, 30 mM ligator, 200 mM imidazole nitrate buffer (pH 6.2), 10 mM MnCl₂

and 100 mM NCI 8. Ligation conditions were based on those reported previously for the conversion of A3'P into A>P (ref. 35) and for the ligation of oligomers with 5P and 2,3-diol termini.

First non-enzymatic self-replicating system



V. Patzke, G. von Kiedrowski ARKIVOC 2007 293-310

SPREAD – Surface-Promoted Replication and Exponential Amplification of DNA Analogues



V. Patzke, G. von Kiedrowski ARKIVOC 2007 293-310

Dynamic oligonucleotide analogue sequence-specific assembly



M. R. Ghadiri et al. Science 2009, 325, 73-77

Dynamic oligonucleotide analogue sequence-specific assembly



M. R. Ghadiri et al. Science 2009, 325, 73-77